Lipid membrane characterization with second harmonic scattering: surface potentials, ionization, membrane asymmetry and hydration

Thèse n. 7626 2017
présentée le 24 Mars 2017
à la faculté des sciences et techniques de l'ingénieur
Chaire Julia Jacobi de Photomédecine - Laboratoire de biophotonique fondamentale
Programme doctoral en biotechnologie et génie biologique
École Polytechnique Fédérale de Lausanne
pour l'obtention du grade de Docteur ès Sciences
par

CORNELIS LÜTGEBAUCKS

acceptée sur proposition du jury:

Prof. Dominique Pioletti, président du jury Prof. Sylvie Roke, directrice de thèse Prof. Francesco Stellacci, rapporteur Prof. Franz Geiger, rapporteur Prof. Eric Vauthey, rapporteur

Lausanne, EPFL, 2017

Berichte aus der Biophysik

Cornelis Lütgebaucks

Lipid membrane characterization with second harmonic scattering: surface potentials, ionization, membrane asymmetry and hydration

Shaker Verlag Aachen 2017 **Bibliographic information published by the Deutsche Nationalbibliothek** The Deutsche Nationalbibliothek lists this publication in the Deutsche Nationalbibliografie; detailed bibliographic data are available in the Internet at http://dnb.d-nb.de.

Zugl.: École Polytechnique Fédérale de Lausanne, Diss., 2017

Copyright Shaker Verlag 2017 All rights reserved. No part of this publication may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise, without the prior permission of the publishers.

Printed in Germany.

ISBN 978-3-8440-5496-5 ISSN 1439-7897

Shaker Verlag GmbH • P.O. BOX 101818 • D-52018 Aachen Phone: 0049/2407/9596-0 • Telefax: 0049/2407/9596-9

Internet: www.shaker.de • e-mail: info@shaker.de

Acknowledgements

First of all I would like to thank my supervisor Professor Sylvie Roke for giving me the opportunity to work with her and doing cutting-edge research for the last 5 years, during which I gained many new insights. I appreciate all the time she invested and the trust she placed in me. A huge thanks goes also to Grazia Gonella for very helpful discussions, her advices, and spending time in hourlong skype calls.

I would also like to thank the LBP group, past and present members, for discussions, and for creating such a kind atmosphere. Specifically, Carlos Macias-Romero, with whom I shared the 5 years in the group and who taught me a lot about optics, but more importantly became a very good friend. Also Rüdiger Scheu, who helped me getting started by answering questions already during breakfast. In addition, I also would like to thank Halil Okur and Yixing Chen for helpful discussions and proof-reading. With these four I also shared an office and that had to endure all my moods over the years.

Furthermore, a huge "thanks" to all the people, colleagues, and friends that I encountered during my stay and that helped creating awesome memories or just for feeding me: Marie Didier, Evangelia Zdrali, Jan Dedic, Orly Tarun, Nikolay Smolentsev, Filip Kovacic, Igor Nahalka, Volker Zagolla, Zahra Monemhagdoust, Enrico Chinello, Andreas Schmocker, Claudia Rodriguez, Miguel Modestino, Manon Rostykus, Clément Javercaz-Galy, Michael Reinke, Yury Kuzminykh, Christophe Moser, Samantha and Mitchell Anderson, Vitalijs Zubkovs, Mikaël Dumortier, Eric Tremblay, Clara Moldova, Matthew Blackburn, Alina Isakova, Miguel Sison, Angel Padua, and Alix D'Agostino.

Thanks to our secretary Rebecca Veselinov for being always helpful and supportive; especially for the small motivations in form of cakes and advent calendars.

Thanks to my comitee, Prof. Pioletti, Prof. Franz Geiger, Prof. Eric Vauthey, Prof. Francesco Stellacci, for taking the time to read my thesis and giving me feedback on my research. Thanks to Prof. Eric Borguet for giving me permission to use some of his data in my work.

Last, I would like to thank my parents for all the love and support they gave me and all the opportunities they provided that let me to this point.

Lausanne, January 2017

Cornelis Lütgebaucks

Abstract

Membranes, composed of a variety of lipids and other biomolecules, mediate signaling processes between cells and their aqueous environment. To fulfill this function, membranes can vary their composition leaflet-specific and thus alter their surface properties. To fully understand the impact of these processes on the molecular level, it is necessary to develop tools that can access the molecular properties of free-floating model membranes label-free. These tools are ideally surface-specific. In this thesis, we apply the nonlinear optical techniques second harmonic scattering (SHS) and vibrational sum-frequency scattering (SFS) together with electrokinetic measurements to label-free characterize the interfacial properties, hydration structure and surface potentials of liposomes in aqueous solutions.

First, we generalize the nonlinear optical theory to describe the second-order surface response from interfaces with aqueous solutions independent of the ionic strength for reflection, transmission and scattering geometries. We demonstrate that interference effects from oriented water molecules in the bulk aqueous solution alter the probing depth and the expected second-order response at low ionic strengths.

Then, we apply this theory to demonstrate that SHS patterns of liposomes and oil droplets contain all necessary information to extract the absolute surface potential of the respective particles without assuming a model for the interfacial structure. By analyzing scattering patterns that capture the orientational distribution of water around the particles, we find surface potentials of -38 mV for bare oil-droplets and -11 mV for zwitterionic liposomes in water. For anionic liposomes the surface potential varies between -150 mV and -23 mV in solutions containing different amounts of NaCl ranging from ~ 0 mM to 10 mM. These values are remarkably different for solutions to the Gouy-Chapman model considering a fixed surface charge density.

Next, we characterize the hydration and lipid asymmetries in binary mixed membranes using SHS and SFS. The liposomes exhibit hydration asymmetry between the inner and outer leaflet. The lipid number density between the inner and outer leaflet is the same, although geometrical packing arguments would suggest a different density. However, an asymmetric lipid distribution between the leaflets can be induced by fine tuning specific intermolecular interactions between the lipids. This is shown with dipalmitoylphosphoserine and dioleoylphosphocholine mixtures creating a membrane

structure that allows intermolecular H-bonding between the phosphate and amine groups of the lipids.

Finally, we quantify the surface properties of membranes composed of lipids containing phosphoserine and phosphocholine headgroups. Surprisingly, we find a very high degree of counterion condensation on anionic membranes in pure water: only 1% of all lipids are ionized. This indicates a tightly packed layer of ions around the membrane that needs to be considered when modelling the interfacial structure around membranes.

Keywords: membranes, lipids, surface potential, ion condensation, hydration, non-linear optics, light scattering, soft matter, liquid/liquid interfaces, electric double layer

Contents

A	cknov	wledge	ments	i
Al	bstra	ct (Eng	dish/Deutsch)	iii
Li	st of	figures		ix
Li	st of	tables		xi
1	Intr	oducti	on	1
	1.1	Lipid	membranes	1
	1.2	Lipids	3	2
	1.3	State	of the art in membrane research	3
		1.3.1	Membrane models	4
		1.3.2	Hydration of membranes and electrostatic interactions $\ \ldots \ \ldots$	7
	1.4	Secon	nd harmonic and sum-frequency generation	9
		1.4.1	Nonlinear processes	9
	1.5	Conce	eptual illustration of SHS from a sphere	11
	1.6		of the art in nonlinear optical scattering of membranes	13
	1.7	This t	hesis	15
2	Exp	erimeı	ntal Details & Methodology	17
	2.1	SHS: 0	Characterization & Assumptions	18
		2.1.1	SHS setup	18
		2.1.2	Characterization of the fundamental laser beam	19
		2.1.3	Validation of elastic SHG	20
		2.1.4	Polarization sensitivity of the setup	21
		2.1.5	Characterization of the setup alignment	22
		2.1.6	Normalization of the surface second harmonic (SH) data $\ \ldots \ \ldots$	23
		2.1.7	General assumptions for nonlinear optical processes	24
	2.2	Comp	olementary experimental techniques & Methodology	27
		2.2.1	Dynamic light scattering (DLS)	27
		2.2.2	Calculation of the scattering efficiency A	29

Contents

		2.2.3	Calculation of an effective radius	30
		2.2.4	Electrophoretic measurements and the ζ -potential	31
	2.3	Samp	le preparation & Characterization	34
		2.3.1	Lipids	34
		2.3.2	Liposome formation	34
		2.3.3	Determination of total phosphorus content	36
		2.3.4	pH stability of aqueous solutions over time	36
3	Mod	dulatio	n of SHG and SFG intensities by interference	39
	3.1	Introd	luction	40
	3.2	Resul	ts & Discussion	41
		3.2.1	SHG/ SFG in reflection mode	41
		3.2.2	SHG/ SFG in transmission mode	48
		3.2.3	SHG/ SFG in scattering mode	49
		3.2.4	Comparison between reflection, transmission, and scattering	
			experiments	54
	3.3	Concl	dusions	57
4	Ext	raction	of the surface potential	59
	4.1		luction	60
	4.2	Mater	rials & Methods	61
		4.2.1	Chemicals	61
		4.2.2	Cleaning procedures	62
		4.2.3	Sample preparation	62
		4.2.4	Angle-resolved (AR)-SHS measurements	63
4.3 Results & Discussion		Resul	ts & Discussion	63
		4.3.1	Physical contributions and origin of the signal for an AR-SHS	
			experiment	63
		4.3.2	AR-SHS theory for a collinear beam geometry $\ \ldots \ \ldots \ \ldots$	64
		4.3.3	Experimental verification of equations	66
		4.3.4	Implementation: Normalization to the bulk water response	69
		4.3.5	Oil droplets and DOPC liposomes in aqueous solutions	70
		4.3.6	$\chi_{\mathbf{s}}^{(2)}$	71
		4.3.7	DOPS liposomes in aqueous solutions vs. ionic strength	72
		4.3.8	Comparison between the SHS derived values and solutions to the	
			Gouy-Chapman and constant capacitor model	73
	4.4	Concl	dusions	74
5	Lipi	d trans	smembrane asymmetry in liposomes	75
	5 1	Introd	Justion	76

	5.2	Mater	ials & Methods	77		
		5.2.1	Lipids	77		
		5.2.2	SHS measurements and normalization	77		
		5.2.3	Vibrational sum-frequency scattering (SFS)	78		
		5.2.4	SFS spectral fitting	79		
		5.2.5	$Calculation \ of \ the \ orientational \ distribution \ of \ phosphate \ groups$	79		
		5.2.6	Calculation of the degree of asymmetry based on geometrical			
			arguments	80		
		5.2.7	Calculation of the degree of asymmetry from the SFS data	80		
	5.3	Result	s & Discussion	81		
		5.3.1	Single component liposomes	81		
		5.3.2	Liposomes from binary mixtures	84		
		5.3.3	Can lipid-lipid interactions drive transmembrane asymmetry? .	86		
		5.3.4	Quantification of transmembrane asymmetry	87		
	5.4	Concl	usions	91		
6	Ioni	zation	of DOPC:DOPS lipid membranes	93		
	6.1		uction	94		
	6.2		ials & Methods	95		
		6.2.1	Chemicals	95		
		6.2.2	Cleaning procedures	95		
		6.2.3	Liposomes	95		
		6.2.4	SHS	95		
	6.3	Result	s & Discussion	95		
		6.3.1	DOPC:DOPS membranes	95		
		6.3.2	DOPC:cardiolipin mixtures	101		
	6.4	Concl	usions	101		
7	Sum	ımary 8	& Outlook	103		
	7.1		nary	103		
	7.2		ok	105		
8	A	endix		107		
0			$\chi^{(2)}$ transformation	107		
	0.1	ρ · to	transformation	107		
9	List	of Pub	lications	109		
Bi	Bibliography 11					
Cu	Curriculum Vitae 13					