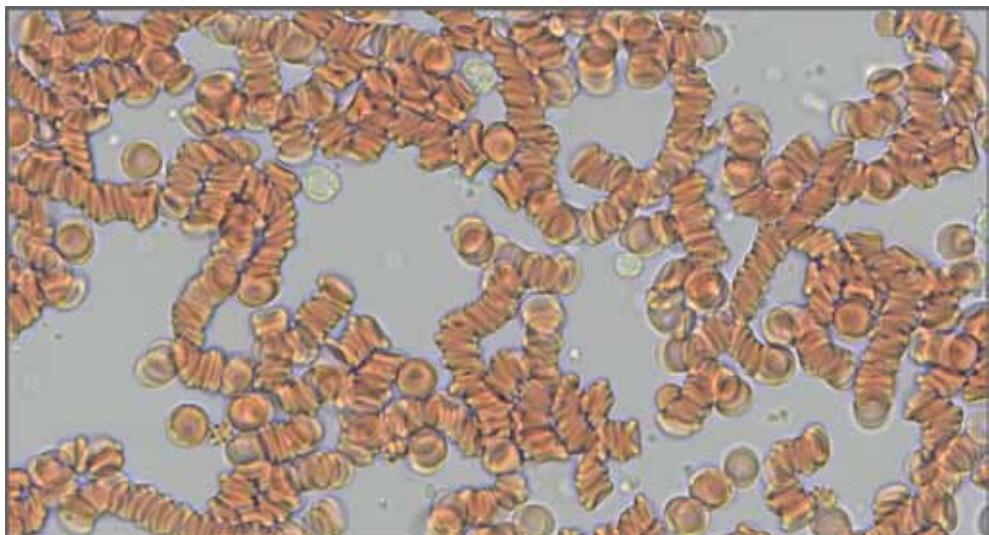


Sebastian V. Jansen

**Development of an In-Vitro
Fluorescent Haemolysis Detection
Method Using Ghost Cells**



Aachener Beiträge zur Medizintechnik

Herausgeber:

Univ.-Prof. Dr.-Ing. Dr. med. Steffen Leonhardt

Univ.-Prof. Dr.-Ing. Klaus Radermacher

Univ.-Prof. Dr. med. Dipl.-Ing. Thomas Schmitz-Rode

Development of an In-Vitro Fluorescent Haemolysis Detection Method Using Ghost Cells

Entwicklung einer fluoreszenzbasierten in-vitro Hämolyse
Detektion mit Ghost Cells

Von der Fakultät für Maschinenwesen der Rheinisch-Westfälischen Technischen Hochschule Aachen zur Erlangung des akademischen Grades eines
Doktors der Ingenieurwissenschaften genehmigte Dissertation

vorgelegt von
Sebastian Victor Jansen

Berichter: Univ.-Prof. Dr.-Ing. U. Steinseifer
 Univ.-Prof. Dr.-Ing. W. Schröder

Tag der mündlichen Prüfung: 07. März 2017

Aachener Beiträge zur Medizintechnik

45

Herausgeber:

Univ.-Prof. Dr.-Ing. Dr. med. Steffen Leonhardt

Univ.-Prof. Dr.-Ing. Klaus Radermacher

Univ.-Prof. Dr. med. Dipl.-Ing. Thomas Schmitz-Rode

Sebastian Victor Jansen

Development of an In-Vitro Fluorescent Haemolysis Detection Method Using Ghost Cells

Ein Beitrag aus dem Institut für Angewandte Medizintechnik der RWTH Aachen, Lehr- und Forschungsgebiet Kardiovaskuläre Technik (Leitung: Univ.-Prof. Dr.-Ing. Ulrich Steinseifer).

**RWTHAACHEN
UNIVERSITY**

Shaker Verlag
Aachen 2018

Bibliographic information published by the Deutsche Nationalbibliothek

The Deutsche Nationalbibliothek lists this publication in the Deutsche Nationalbibliografie; detailed bibliographic data are available in the Internet at <http://dnb.d-nb.de>.

Zugl.: D 82 (Diss. RWTH Aachen University, 2017)

Copyright Shaker Verlag 2018

All rights reserved. No part of this publication may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise, without the prior permission of the publishers.

Printed in Germany.

ISBN 978-3-8440-5704-1

ISSN 1866-5349

Shaker Verlag GmbH • P.O. BOX 101818 • D-52018 Aachen

Phone: 0049/2407/9596-0 • Telefax: 0049/2407/9596-9

Internet: www.shaker.de • e-mail: info@shaker.de

**To My Parents
and
Miriam**



Acknowledgements

I would like to express my great appreciation to Univ.-Prof. Dr.-Ing. Ulrich Steinseifer, as my advisor, for his scientific guidance, his encouragement and the numerous fruitful discussions during the course of this work at the Department of Cardiovascular Engineering of the Institute of Applied Medical Engineering, RWTH Aachen University. I also want to thank Univ.-Prof. Dr.-Ing. Wolfgang Schröder for co-reviewing this thesis.

I am deeply grateful for my good friend and colleague Jan Roggenkamp, for the innumerable hours of conversations and his great support, also on non-work-related problems.

My sincere thanks go to Max Nachtsheim and Indra Müller, and to all my other student workers who helped me realizing the haemolysis detection method. Special thanks go to the technical staff and the mechanical workshop for making this work possible. Here, I would like to mention Ilona Mager and Judith Maas in particular, for their expertise and assistance during the innumerable blood trials. Great thanks to all my colleagues for creating a remarkable working environment and atmosphere that accounted for numerous productive and enriching discussions during my work and resulted in some very good friendships including Stephanie Schmitz, Sabrina Herren, Melina Krämer, Kathrin Gester, Martin Büsen, Felix Hesselmann, Johanna Clauser and many more.

I deeply want to thank Dorothee Roggenkamp, Dr. Sebastian Burgmann and Dr. Sebastian Große for their constant advise and support as well as for the enlightening discussions, usually on the beach in Domburg.

My education is considerably based on the work and sacrifices of my parents who always supported and encouraged me throughout my life in order to expand my curiosity and to find my own way. It is hard to express my gratitude towards them in just a few lines. I am also deeply indebted to my wife Miriam Jansen for her continuous support, humour and love that makes me looking forward into the adventures of the upcoming life. To her and to my parents, I dedicate this thesis.

Abstract

Cardiovascular devices that are in direct contact to the blood stream, such as heart-valve prostheses or blood pumps, need to be designed with respect to a minimal extent of blood damage. Device induced, flow field related stress can cause mechanical destruction of blood cells (haemolysis) or artificially trigger a physiological blood clotting reaction (thrombosis) with life-threatening consequences. In order to analyse the risk of blood damage in a device, a variety of experimental and numerical methods are available. However, experimental methods are currently impeded by the inapplicability of optical measurement techniques, due to the low transparency of blood, which is majorly determined by the protein haemoglobin inside the red blood cells. The haemoglobin content of the red blood cells can be diluted by a special procedure and the remaining haemoglobin-poor erythrocytes are termed ghost cells. Due to the reduced haemoglobin content, ghost cells possess a markedly higher transparency compared to normal erythrocytes.

This thesis shows that a suspension of ghost cells can be used as a blood analogue fluid in combination with optical measurement techniques in order to improve the determination of blood damage. Especially, a novel optical measurement principle based on ghost cells is presented that is able to detect haemolysis spatially resolved for the first time. Ghost cells are prepared from porcine erythrocytes using a customized procedure. The rheology of the ghost cell suspension is comparable to normal blood, whereas the transparency of the suspension is highly increased and enables the use of optical measurement techniques. The novel haemolysis detection method, for which the term Fluorescent Haemolysis Detection (FHD) is proposed, is based on the idea that a target substance is enclosed in the ghost cells and a corresponding fluorescent indicator is suspended in the plasma. In the case of a cell lysis (haemolysis) the indicator binds to the target and emits a fluorescent signal which can be recorded by a camera. A corresponding experiment proofs the functioning of the principle. This might help manufacturers of cardiovascular devices to reduce device-induced blood trauma and might boost the understanding of the underlying mechanisms of haemolysis and thrombosis.

Übersicht

Kardiovaskuläre Systeme in direktem Blutstrom, wie zum Beispiel Herzklappenprothesen oder Blutpumpen, müssen im Hinblick auf eine minimale systeminduzierte Blutschädigung ausgelegt werden. Das vom System veränderte Strömungsfeld und dessen Kräfte können zur mechanischen Zerstörung von Blutzellen (Hämolyse) führen sowie eine Gerinnungsreaktion auslösen (Thrombose) mit möglicherweise lebensgefährlichen Konsequenzen. Es existieren eine Reihe von numerischen und experimentellen Methoden, um die Blutschädigung eines Systems zu analysieren. Eine Limitation dieser Methoden ist jedoch die mangelnde Transparenz von Blut, die eine Anwendung optischer Methoden nicht möglich macht. Die Opazität des Bluts wird durch das Protein Hämoglobin in den Erythrozyten hervorgerufen. Durch eine Prozedur kann der Hämoglobingehalt der Erythrozyten verringert werden. Die hämoglobinarmen Erythrozyten werden Ghost cells genannt und sind wesentlich transparenter als normale Erythrozyten.

In dieser Doktorarbeit wird gezeigt, dass eine Suspension mit Ghost cells als blutanaaloges Fluid für optische Messmethoden eingesetzt werden kann und somit die Analyse von Blutschädigung verbessert. Insbesondere wird ein neuartiges Messprinzip vorgestellt, welches zum ersten Mal eine ortsaufgelöste Detektion von Hämolyse ermöglicht. Ghost cells werden aus Erythrozyten von Schweineblut in einer angepassten Prozedur hergestellt. Die Rheologie der Suspension ist vergleichbar mit normalem Blut, die Transparenz ist dagegen deutlich höher und ermöglicht die Anwendung optischer Messmethoden. Die neuartige Hämolsedetektion basiert auf der Idee eine Targetsubstanz in den Ghost cells einzuschließen, wohingegen ein fluoreszierender Indikator ins Plasma suspendiert wird. Im Falle der Zellyse (Hämolyse) bindet der Indikator an das Target und emittiert ein Fluoreszenzsignal, welches mit einer Kamera aufgezeichnet werden kann. Die Funktion dieses Prinzips wird in einem entsprechenden Experiment gezeigt. Dieses Prinzip kann Herstellern kardiovaskulärer Systeme bei der Reduzierung systeminduzierter Blutschädigung helfen. Darüber hinaus kann eine solche Methode auch zu einem größeren Verständnis über die zugrundeliegenden Mechanismen von Hämolyse und Thrombose beitragen.

Contents

Abstract.....	III
Nomenclature.....	VII
1 Introduction.....	1
2 Flow Induced Blood Damage	5
2.1 Blood Flow.....	7
2.1.1 Rheological Properties of Blood	8
2.1.2 Blood Flow Measurement Using Particle Image Velocimetry.....	12
2.1.3 Other Blood Flow Measurement Techniques	14
2.2 Haemolysis	16
2.2.1 Experimental Haemolysis Measurement Techniques.....	18
2.2.2 Numerical Haemolysis Models.....	19
2.3 Thrombosis	21
2.3.1 Basic Principle of Haemostasis	21
2.3.2 Flow Induced Thrombus Formation	26
2.3.3 Experimental Thrombus Measurements.....	28
2.3.4 Numerical Models of Thrombosis.....	31
3 Production and Properties of Ghost Cells.....	35
3.1 Porcine vs. Human Blood.....	36
3.2 Erythrocyte Membrane Structure	37
3.3 Production of Ghost Cells.....	39
3.3.1 Parameter Setup	40
3.3.2 Utilized Blood and Solutions	42
3.3.3 Production Procedure	44
3.4 Properties of Ghost Cells.....	48
3.4.1 Size and Shape of Ghost Cells.....	48
3.4.2 Optical Properties of Ghost Cells	55
3.4.3 Density	59
3.4.4 Rheology	63
3.5 Large Scale Production	66
3.5.1 Filter Apparatus	67
3.5.2 Centrifuge vs. Filter Process.....	72

4 Fluorescent Haemolysis Detection (FHD).....	81
4.1 General Principle	81
4.2 Preparation of FHD Ghost Cell Suspension	83
4.2.1 Target and Ghost Cells	83
4.2.2 Fluorescent Indicator and Artificial Plasma.....	84
4.3 Proof-of-Principle.....	87
4.3.1 Experimental Setup	88
4.3.2 Results	89
4.3.3 Discussion and Conclusion	92
5 Further Applications of Ghost Cells	95
5.1 PIV Measurements Using Ghost Cells.....	95
5.1.1 Experimental Setup	96
5.1.2 Results	98
5.1.3 Discussion and Conclusion	99
5.2 Fluorescent Thrombus Detection (An Outlook).....	101
5.2.1 Coagulability of Ghost Cell Suspensions.....	102
5.2.2 Further Outlook	106
6 Summary and Conclusion.....	107
References	111
Appendix.....	129